preliminary notes and applications from Bioanalytical Systems, Inc.

# Erythromycin in Neonatal Serum

## **Purpose**

Determination of erythromycin in neonatal serum.

$$H_3$$
C  $CH_3$   $CH_3$ 

Figure 1. Structures of enythromycin [1] and roxithromycin [1].

[ii] R = N-OCH2-OCH2CH2OCH3

Erythromycin is a macrolide antibiotic frequently used for penicillin-resistant bacterial and mycoplasmic infections. In particular, it is being tested for the treatment of chronic lung disease of premature neonates, which may be caused by *Ureaplasma urealyticum* [1].

Sample sizes for these patients range from 10 to 100  $\mu$ L. Therefore, it was necessary to develop a highly sensitive assay [2]. Moreover, since other antibiotics are commonly co-administered, the assay had to be highly selective.

Although not ideal electrophores, the tertiary amino groups often present in macrolide antibiotics are capable of anodic oxidation under sufficiently basic pH conditions [3]. Other antibiotics in this class that contain an aliphatic tertiary amine on the desosamine ring (clarithromycin, roxithromycin, azithromycin, dirithromycin) should have similar electrochemical properties.

## **Existing Method**

LCUV, and LCEC at more neutral pH. Neither has the sensitivity required for the small samples obtained from neonates.

#### **Conditions**

System: BAS-200 Liquid Chromatograph Detector: Dual-series, glassy carbon

Potential: Upstream: +1500 mV; Downstream: +875

mV (both vs. Ag/AgCI)

Column: Chromegabond Gamma RP1, 150 x 4.6 mm, 5 µm particle size (ES Industries, Marlton NJ) Mobile Phase: 160 mL methanol, 60 mL n-propanol, 780 mL 0.016 M potassium phosphate/0.04 M sodium perchlorate (pH 11).

Flow Rate: 1 mL/min Temperature: Ambient Detection Limit: 32 ng/mL Linear Range: 0.03 - 6 µg/mL

#### Sample Preparation

To 100  $\mu$ L samples, add 50  $\mu$ L roxithromycin (12  $\mu$ g/mL, Internal Standard) and vortex. Add 200  $\mu$ L 0.06 M Na<sub>2</sub>CO<sub>3</sub> and vortex. Add 1 mL methyl-*t*-butyl ether. Vortex, centrifuge, and transfer the ether layer to a new tube. Evaporate to dryness and reconstitute in 63  $\mu$ L of mobile phase which has been adjusted to pH 8. Inject 55  $\mu$ L.

## **Notes**

Only the downstream electrode was monitored. Setting a high potential on the upstream electrode stabilized the baseline obtained from the downstream electrode.

F2 and F3 are chromatograms of serum from a patient, shortly after an initial dose and 24 hours after the final dose.

Pharmacokinetic data are presented in F4 and F5.

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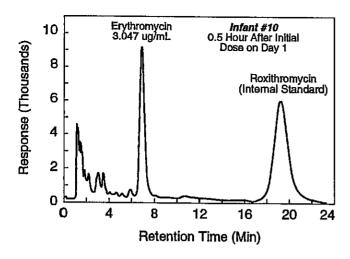


Figure 2. Sample from one patient shortly after the initial dose.

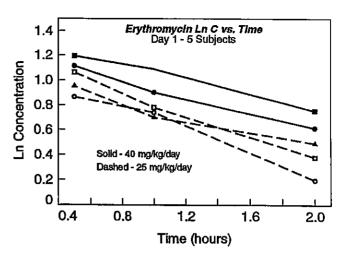


Figure 4. Two-hour pharmacokinetic data.

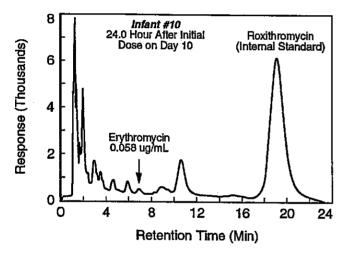


Figure 3. Sample from the same patient a day after the final dose.

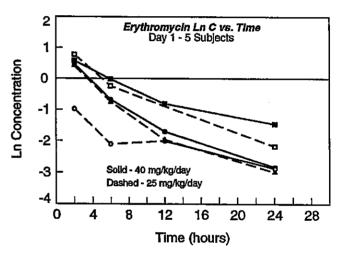


Figure 5. Twenty-four hour pharmacokinetic data.

## References

- 1. G.H. Cassell, K.B. Waites and D.T. Crouse, Clinics in Perinatology 18 (1991): 241-262.
- R.E. Shoup, S. Hessong, K.B. Waites and D.W. Knuth, poster session presented at the American Association of Pharmaceutical Scientists 6<sup>th</sup> Annual Meeting, Washington. D.C., 1991.
- 3. G.S. Duthu, J. Liquid Chromatogr. 7 (1984): 1023.

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